REVIEW



Conjugated linoleic acid and glucosamine supplements may prevent bone loss in aging by regulating the RANKL/RANK/OPG pathway

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Abstract

The skeleton is a living organ that undergoes constant changes, including bone formation and resorption. It is affected by various diseases, such as osteoporosis, osteopenia, and osteomalacia. Nowadays, several methods are applied to protect bone health, including the use of hormonal and non-hormonal medications and supplements. However, certain drugs like glucocorticoids, thiazolidinediones, heparin, anticonvulsants, chemotherapy, and proton pump inhibitors can endanger bone health and cause bone loss. New studies are exploring the use of supplements, such as conjugated linoleic acid (CLA) and glucosamine, with fewer side effects during treatment. Various mechanisms have been proposed for the effects of CLA and glucosamine on bone structure, both direct and indirect. One mechanism that deserves special attention is the regulatory effect of RANKL/RANK/OPG on bone turnover. The RANKL/RANK/OPG pathway is considered a motive for osteoclast maturation and bone resorption. The cytokine system, consisting of the receptor activator of the nuclear factor (NF)-kB ligand (RANKL), its receptor RANK, and its decoy receptor, osteoprotegerin (OPG), plays a vital role in bone turnover. Over the past few years, researchers have observed the impact of CLA and glucosamine on the RANKL/RANK/OPG mechanism of bone turnover. However, no comprehensive study has been published on these supplements and their mechanism. To address this gap in knowledge, we have critically reviewed their potential effects. This review aims to assist in developing efficient treatment strategies and focusing future studies on these supplements.

Keywords Aging · Conjugated linoleic acid · Glucosamine · Bone loss · RANKL/RANK/OPG pathway

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Introduction

The skeleton is a living organ that is in a permanent state of turnover, including bone resorption and formation [1]. The adult human skeleton comprises approximately 206 bones [2]. Bones are responsible for body form and protecting vital organs, such as the brain and heart. Additionally, there are stem cells in the bone marrow that are remarkably important for hematopoiesis [3, 4]. As the world population ages and life expectancy increases, the number of people who suffer from bone discomfort also increases. According to reports, more than 200 million individuals worldwide struggle with osteoporosis. The International Osteoporosis Foundation indicates that one in three women and one in five men experience bone discomfort in their lifetime [5]. Bone mass density is a value used to measure bone health and strength, which naturally decreases due to aging or pathologically due to some diseases. Losing about 0.5-1% of bone mass each year after achieving peak bone mass is a

natural occurrence. Recent investigations indicate that men and women are at risk of bone loss in almost equal amounts, but the difference is that women usually suffer from severe bone loss during postmenopause and afterward, while bone resorption in men occurs slowly over time [6]. Alongside genetic factors, physical activity, and a healthy food regime affect bone health. Various materials are needed for a healthy bone structure, such as calcium, magnesium, and phosphate [7], as well as protein and fat [8]. Nowadays, various methods are used to protect bone health, including drugs, surgeries, and physical therapies [9]. Bisphosphonates [10], Denosumab, Calcitonin [11], and Estrogen antagonists [12] are some of the most common medications used to prevent bone loss. Nutrient factors like vitamin D, vitamin K, and calcium [13], whether obtained through diet or supplements, can also be useful for bone amplification.

On the other hand, several medications can put bone health at risk and lead to bone loss. These medications include glucocorticoids, thiazolidinediones, heparin, anticonvulsants, chemotherapies, and proton pump inhibitors [14].

Osteoblasts [15], osteoclasts [16], osteocytes [15], and bone lining cells are the fundamental cells of bones. In a healthy body, the balance between osteoblasts (responsible for bone formation) and osteoclasts (responsible for bone resorption) keeps the skeleton healthy [17]. Recent investigations have been focusing on strategies to enhance bone formation [18–21]. In the middle of 1990s, it was revealed that osteoclast maturation depends on three markers belonging to the TNF superfamily. Receptor activator of NF-kB (RANK) is a receptor on the surface of preosteoclasts. Its binding with receptor activator of NF-kB ligand (RANKL) marks the onset of preosteoclast changes. Osteoprotegerin (OPG) participates as a decoy receptor for RANKL and prevents RANKL/RANK interaction. In conclusion, a higher OPG/RANKL ratio is a symptom of lower bone resorption [22].

The RANKL/RANK/OPG pathway is a crucial cytokine system involved in the regulation of bone turnover, and its significance in bone health has been extensively documented in recent papers [22, 23]. This pathway plays a pivotal role in osteoclast maturation and bone resorption, making it a fundamental mechanism to investigate in the context of bone health [22, 24]. The RANKL/RANK/OPG system stands as one of the most significant discoveries in bone biology in the past decade, and its disruption leads to or causes numerous bone diseases. Various factors, including cytokines, hormones, and growth factors, regulate the RANKL/RANK/OPG pathway [23]. In conclusion, a higher OPG/RANKL ratio is a symptom of lower bone resorption, and the elimination of RANKL and RANK in animal studies shows a major effect in inhibiting bone mass loss and osteoporosis [22, 23].

Conjugated linoleic acid (CLA) is a type of polyunsaturated fatty acid that contains two double bonds between carbons in conjugated and is made by an isomerization pathway [25]. There are two major forms of CLA: cis-9 trans-11 CLA and trans-10 cis-12 CLA [26]. Many advantages have been suggested for CLA, including anti-carcinoma effects, anti-diabetes effects, and a reduced risk of atherosclerosis [27]. Athletes have used CLA specifically for its effect of reducing body fat mass and boosting lean body mass to maximize exercise outcomes [28]. Furthermore, some examinations have shown that CLA's anti-inflammatory effects could be beneficial in reducing post-exercise inflammation in athletes [29].

Glucosamine is an essential precursor for glycoproteins and glycosaminoglycans. It is a glucose-derived amino sugar that can be naturally produced in the human body [30]. Apart from its natural role in cartilage and synovial fluid, glucosamine could have anti-inflammatory effects as a supplement [31]. Additionally, glucosamine has been used to repair joint injuries and alleviate skeletal pain in athletes. Exercise-induced upregulation in athletes' cartilage metabolism increases the need for glucosamine as well [32].

CLA and glucosamine are under investigation in new studies as potential therapeutic approaches with fewer side effects.

The selection of conjugated linoleic acid (CLA) and glucosamine supplements was based on their potential benefits for bone health, as supported by existing research [32, 33]. CLA has been shown to have positive effects on bone structure in various in vitro and in vivo studies. It has been found to increase bone mass by reducing bone resorption and increasing bone mineralization [6]. Glucosamine, on the other hand, has shown promise in supporting healthy bone growth and reducing the weakening of bones. Early studies suggest that glucosamine supplements may help prevent the progression of osteoporosis post-menopause [34]. The direct and indirect effects of CLA and glucosamine on bone structure have been linked to a number of different mechanisms. One particular mechanism that demands special focus is the regulatory impact of RANKL/RANK/ OPG on bone turnover. This mechanism is common in bone turnover effects of both CLA and glucosamine [35, 36]. The RANKL/RANK/OPG pathway is considered a motive for osteoclast maturation and bone resorption [24]. However, no in-depth research on these supplements, particularly the RANKL/RANK/OPG pathway, has been released. We have critically examined their potential effects to fill in this knowledge gap, guide future research on these supplements, and aid in the development of effective treatment plans.

Two common sports supplements: conjugated linoleic Acid/Glucosamine and bone health

Conjugated linoleic acid

CLA is an isoform of linoleic acid with a conjugated double bond. The term "conjugated" refers to molecules having a single bond separating two double or triple bonds. Conjugated linoleic acids are 18-carbon fatty acids (C18:2 n-6), mostly having 2 double bonds in a conjugated form at positions (9, 11) or (10, 12) [37]. The amount of CLA humans receive from dietary sources is about 10% of the suggested CLA per day. The content of CLA intake from dietary sources in the USA is about 104-151 mg in women and 176-212 mg in men [38]. Based on animal studies, at least 3 g/d of CLA is recommended to achieve its health benefits [39]. Almost 4/5 of the existing CLA in dietary sources is c9t11-CLA. Most CLA supplements contain the same amount of CLA isomers [40]. Synthetic forms of CLA are usually plant-based and typically contain equal amounts of cis 9, trans 11, and trans 10, cis 12 [41].

Dietary sources of CLA, including meats, eggs, and dairy, contain cis-9, trans-11, and trans-10, cis-12 in a 3:1 ratio [40]. Animal age, diet, and species are some of the factors influencing the amount of CLA in their tissues or products, such as milk or eggs [42]. For example, feeding animals with oilseeds and fish oil could result in more CLA production in their tissues [43].

Multiple microorganisms have the ability to perform CLA biohydrogenation [44]. In fact, CLAs are intermediates formed during the production of polyunsaturated fatty acids such as linoleic acid and linolenic acid [43]. Several investigations have shown that CLA is the product of a species of *Lactobacillus acidophilus*. These microorganisms produce CLA from linoleic acid under microaerobic conditions [44]. Hydroxy fatty acids like 10-hydroxy-12-octadecenoic acid are intermediates in the CLA production pathway [45]. *Butyrivibrio fibrisolvens* is another ruminal microorganism participating in CLA production in animal tissues [46]. Additionally, *Clostridium sporogenes, Bifidobacterium breve, Propionibacterium acnes, Lactobacillus Reuteri*, and *Propionibacterium freudenreichii* are named as bacterial agents of CLA production [47].

In a mice study, it was reported that CLA could prevent chemically-induced skin neoplasia, leading to comprehensive studies about CLA's health benefits [48]. Based on studies, several effects have been suggested for CLA intake. It has been shown that CLA intake could lower the risk of type 2 diabetes, cancer, and atherosclerosis, and it might be helpful for weight loss [44]. It seems that the anti-carcinogenic and metabolism-regulation effects of CLA are influenced by modification in PPAR α and γ [49]. It has been reported that t10c12 CLA appears to suppress lipid uptake by blocking lipoprotein lipase and stearoyl-CoA desaturase activities. Additionally, it has been shown to regulate lipid metabolism in hepatic cells in in-vitro studies [50]. Furthermore, several studies suggest that CLA has advantages for diabetic people, as it reduces insulin resistance and increases GLUT4 expression [51]. Regarding CLA's anti-atherosclerosis effects, various mechanisms have been suggested, including reducing macrophage LDL uptake, monocyte adhesion, and monocyte migration, all leading to the improvement of the damaged environment and regulating atherosclerotic mediators [52]. However, several investigations have achieved some undesirable effects of CLA consumption, such as oxidative stress, insulin resistance, and gastrointestinal irritation [27]. These contradictory results indicate the importance of more studies about CLA consumption, especially CLA supplementation.

CLA's effect on bone metabolism is one of the most ambiguous effects of CLA on the human body. The limited information available suggests that CLA affects the skeleton mainly by reducing prostaglandins, which could augment bone resorption and formation based on their concentration in bones, and increasing calcium absorption [53]. Several animal and in vitro studies have shown that CLA supplementation leads to a lower risk of bone fractures by different mechanisms (Table 1) [41].

Glucosamine

Investigations are examining new alternative strategies to control the degeneration of cartilage in bone-related diseases, including osteoarthritis [54-56]. Glucosamine (2-amino-2-deoxy-β-d-glucopyranose), as a possible treatment agent, is a glucose-based endogenous amino-monosaccharide primarily expressed in cartilage and connective tissues [57]. Its active form, D-glucosamine, results from an interaction between fructose 6-phosphate and glutamine through the hexosamine biosynthesis pathway [58]. Glucosamine appears to participate in cartilage and synovial fluid as part of glycosaminoglycan, proteoglycan, and hyaluronic acid structures [59]. Moreover, some evidence indicates that glucosamine has anti-inflammatory, antiviral, and antioxidative effects [30]. Various studies suggest that glucosamine could inhibit inflammation by suppressing the Nf-Kb pathway [58]. New strategies of dietary herbal supplements focus on the regulation of inflammation and immune regulation as target mechanisms [60–63].

In addition to endogenous production, glucosamine supplements are used as adjuvant therapy for several health issues, including osteoarthritis [64]. Since there are few food sources of glucosamine, most supplements are obtained

studyyear participants/species treatment/dosage/duration significant outcomes Rahman et al., RAW2647-Fells, a CLA e9t11, CLA t10c12 The RANKL-induced NF-e8 activation, including 1-B phosphorylation, 1-B degradation, murrine monocytic cell 2006 murrine monocytic cell 9,11 and 10,12 (> 98% pure) isomes Despite not having a substantial impact on osteoclast formation, 10,12 CLA decreased ost ince 2011 0,11 - CLA, 9,5% CO+0.5% and L-6 osteoclastogenic factors. Despite not having a substantial impact on osteoclast formation, 10,12 CLA decreased ost clast activity, enthepsin K, and RANK expression. 2011 mice CLA, 9,5% CO+0.5% and L-6 osteoclastogenic factors. 2011 mice CLA siomers (9,5% CO+0.5% and L-6 osteoclastogenic factors. 2011 mice CLA siomers (9,5% CO+0.5% and L-6 osteoclastogenic factors. 2011 mice CLA siomers (9,5% CO+0.5% and L-6 osteoclastogenic factors. 2011 mice CLA siomers 9,5% CO+0.5% 2011 mice CLA siomers 9,5% CO+0.5% 2011 mice CLA siomers 10,1612-CLA) for 2011 model of collagemase glucosamine at advector (1	Table 1 Summary	of research that were perf	cormed to assess the effects of CLA/glucc	Table 1 Summary of research that were performed to assess the effects of CLA/glucosamine on bone related RANKL/RANK/OPG pathway	
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model of collagenase- induced osteoarthritisglucosamine at a dose of 20 mg/kg/ daily for 20 days1induced osteoarthritis mouse osteoblasticdaily for 20 daysmouse osteoblasticGlucosamine and N-acctyl-D-glucosamineMC3T3-E1 cellsN-acctyl-D-glucosaminemice0.5% CLA for 24 weeksmice0.5% CLA for 24 weekshuman OA subchondral bone osteoblastsCS (200 µg/mL), glucosamine (50 and 200 µg/mL)patients with kneeglucosamine sulfate plus etoricoxib osteoarthritis	nan et al.,	mice	CLA isomers (9.5% CO + 0.5% c9t11-CLA, 9.5% CO + 0.5% t10c12-CLA or 9.5% CO + 0.25% c9t11-CLA + 0.25% t10c12-CLA) for 6 months	a rise in BMD followed by a reduction in the serum levels of RANKL, TRAP5b, TNF-alpha, and IL-6 osteoclastogenic factors.	[36]
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human OA subchondral CS (200 μg/mL), glucosamine (50 bone osteoblasts and 200 μg/mL) patients with knee glucosamine sulfate plus etoricoxib osteoarthritis		mice	0.5% CLA for 24 weeks	The expression of osteoprotegerin, a RANKL decoy receptor, was activated by CLA while RANKL was inhibited. CLA decreased pro-inflammatory cytokine and raised anti-inflammatory cytokine concentration of lipopolysaccharide-stimulated splenocytes and BM cells.	[88]
patients with knee glucosamine sulfate plus etoricoxib osteoarthritis	Tat et al., 2007	human OA subchondral bone osteoblasts	CS (200 μ g/mL), glucosamine (50 and 200 μ g/mL)	Glucosamine decreased the ratio of OPG/RANKL and resorption activity.	[35]
		patients with knee osteoarthritis	glucosamine sulfate plus etoricoxib	The intevention decreased CTX-II, COMP, and RANKL levels.	[91]

from shrimp shells, chitosan, and other sea crustaceans [65]. Glucosamine supplements are reportedly safe and have no serious side effects, but they might cause some general side effects, such as gastric pain and diarrhea. Additionally, allergic reactions might occur due to the sea-crustacean-based ingredients [66].

Based on different studies, oral consumption of glucosamine with 26% bioavailability is the most common way of taking glucosamine. Glucosamine sulfate and glucosamine hydrochloride are the most popular types of glucosamine supplements [30]. Valid evidence shows that glucosamine sulfate has the highest bioavailability among all forms of glucosamine supplements [67].

CLA and bone health

Various investigations have examined the effects of CLA on bone health, both in vivo and in vitro. Since most of the studies on CLA's skeletal effects are animal studies, their results partly depend on the type of animal being examined. Investigations have shown that CLA consumption might increase bone mineral density in chicks and mice, but no significant changes were observed in pigs and rats [68].

A number of mechanisms have been proposed for CLA's effects on bone structure, both direct and indirect (see Fig. 1). Consuming dietary CLA can increase bone formation by regulating prostaglandin E2 in young chicks [36]. In human osteoblast-like cells, PGE2 is reduced with the consumption of trans-10, cis-12 CLA, but no effects were observed from cis-9, trans-11. PGE2 has been described as a modulator for bone formation and resorption [41]. The reduction in PGE2 might result from arachidonic acid restriction [49]. Additionally, an in vitro study has suggested CLA's potential to upregulate alkaline phosphatase activities [69]. Alkaline phosphatase is an enzyme that plays a major role in bone mineralization through the hydrolysis of phosphatecontaining strata, synthesizing orthophosphate, and enhancing calcium uptake [70]. Despite its benefits, CLA isomers could have some unwanted effects on the skeleton. Some experiments revealed that the 9 cis, 11 trans isomer of CLA resulted in the growth of the number and size of mineralized bone nodules [71].

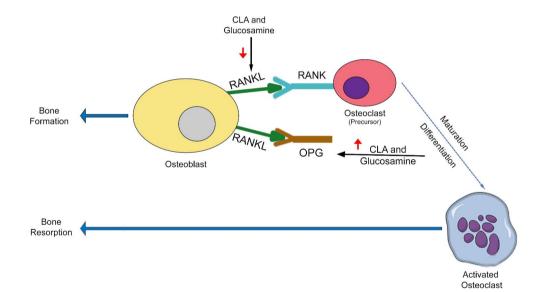


Fig. 1 An illustration of the RANKL-RANK-OPG pathway. Numerous cell types, including osteoblasts and bone marrow stromal cells, produce RANKL as a membrane-bound or secreted ligand. When RANKL binds to the corresponding RANK receptor on the surface of these cells, a number of intracellular processes are triggered, resulting in the maturation and differentiation of precursor osteoclasts into activated osteoclasts. Possible blocking RANKL expression by CLA and glucosamine leads to diminished levels of the RANKL/RANK combination, which in turn prevents the maturation of the precursor of osteoclast to activated osteoclast. The RANKL/RANK signaling pathway plays a crucial role in osteoclast formation, activation, and survival. When RANKL binds to its receptor RANK on the surface of osteoclast precursor cells, it triggers the fusion and differentiation of these cells into mature osteoclasts. However, blocking the interaction between RANKL and RANK can inhibit osteoclastogenesis and prevent the formation of mature osteoclasts. This can be achieved by

using specific inhibitors or agents including CLA and glucosamine that target RANKL expression or interfere with the RANKL-RANK interaction. By disrupting this pathway, the maturation of osteoclast precursors into activated osteoclasts can be effectively prevented, thereby potentially reducing bone resorption and maintaining bone health. The soluble RANKL decoy receptor, OPG, which shares structural similarities with RANK, can inhibit the activity of RANKL. By blocking RANKL's ability to associate with RANK, OPG prevents all downstream molecular processes that would otherwise result in osteoclast differentiation and bone resorption. CLA and glucosamine can regulate bone remodeling and osteoclast activation by increasing OPG expression and decreasing RANKL in osteoblasts. Therefore, by increasing OPG expression and decreasing RANKL, CLA and glucosamine can affect bone resorption by decreasing mature osteoclast formation and activity. Abbreviations: RANKL, receptor activator of nuclear factor (NF)-kB ligand; OPG, osteoprotegrin; CLA, conjugated linoleic acid

It seems that feeding animals with CLA-rich food like butter leads to an increase in bone synthesis [72]. Another experiment has indicated that simultaneous use of CLA and calcium may possibly protect the bone against postmenopausal bone loss [73]. Co-supplementation of CLA and calcium modulated bone markers and caused a slowing down in bone resorption [73]. A closer look indicated that CLA supplementation could augment bone mineral resources in animals with higher calcium in their diet [68]. Some investigations suggest that regulating PPAR γ leads to an enhancement in osteoblasts' number and bone formation after a bone fracture. As CLA could modulate PPAR, CLA is suggested for bone treatment [49].

Glucosamine and bone health

Glucosamine supplementation has widespread use in bonerelated disorders, not only as an over-the-counter dietary supplement (in the US) but also as a slow-acting medication (in Europe), particularly in osteoarthritis (OA) [65]. Several investigations have indicated that 1500 mg/day of glucosamine sulfate improved OA symptoms in many cases. However, glucosamine hydrochloride has not shown the same impact [74]. Moreover, the anti-inflammatory effect of glucosamine sulfate is comparable to NSAIDs such as ibuprofen, with better toleration in patients [74]. Limited trials that have been done on glucosamine's effects in osteoporosis support the idea of its anti-osteoporotic effects as well [75]. It is indicated that the mechanism of glucosamine supplements is basically the same as the endogenous form of it [66]. In vitro and in vivo investigations have suggested different mechanisms of action for glucosamine in bone health, including regulating glycosaminoglycans and type II collagen expression, reducing inflammatory agents, down-regulating catabolic enzymes, and interfering with the NF-kB pathway (Table 1) [76].

As mentioned above, there seem to be potential effects of CLA and glucosamine supplements on bone health. Although various inflammatory mechanisms may be involved in the effective role of these supplements, it seems that they can exert their protective effects by regulating the RANKL/RANK/OPG pathway.

RANKL/RANK/OPG Pathway

The discovery of the RANKL/RANK/OPG pathway was the result of long and accurate investigations aimed at finding the mechanisms of osteoclast maturation and bone resorption [77]. The cytokine system formed by receptor activator of nuclear factor (NF)-kB ligand (RANKL), its receptor RANK, and its decoy receptor Osteoprotegerin (OPG) has a

vital role in bone turnover [78]. Additionally, this pathway has a role in the alteration of breast glands through pregnancy and progesterone-positive breast cancer [79]. It has been suggested that the RANKL/RANK/OPG system is an effective factor in autoimmune diseases such as rheumatoid arthritis and might have interactions with the immune system as well [80].

RANKL (receptor activator of NF-kB ligand) is a transmembrane protein expressed by osteoblasts, tumor cells, and immune cells. As a ligand, RANKL starts a chain reaction in immature osteoclasts when it binds to its receptor, RANK, a TNF receptor superfamily member expressed on the osteoclasts' membrane. The RANKL-RANK connection starts a cascade of reactions in osteoclasts that result in osteoclast maturation [81].

Osteoprotegerin (OPG), another member of the TNFreceptor superfamily, which, unlike RANK, is not bonded to the cell membrane surface, acts as a decoy receptor for RANKL. OPG, produced by osteoblast lineage cells and several other cells in the bone marrow, binds with RANKL to prevent RANKL/RANK connection and following reactions. As a result, the number of mature osteoclasts naturally decreases, and bone resorption is controlled [82]. Based on former findings, when some mediator decreases RANKL expression level, most of the time, OPG expression gets somewhat limited or even decreases as well [22]. Additionally, OPG expression is affected by cytokines, hormones, growth factors, and Wnt/ β -catenin [22].

In most bone diseases, many agents like hormones, cytokines, and growth factors play their roles by making changes in OPG levels, RANKL levels, and disrupting their balance [83]. An enhancement in the RANKL/OPG ratio is observable in metastatic bones. This condition could put bone integrity in danger, lead to some unwanted changes, and also keep the risk of tumor returns at high levels [84]. Bone discomforts, especially osteoporosis, usually have a significant relation to individuals' genetics. Several studies reported that single nucleotide polymorphisms (SNPs) located near TNFRSF11B, TNFSF11, and TNFRSF11A are in charge of encoding OPG, RANKL, and RANK, respectively [85].

Although RANKL, RANK, and OPG have the main roles in this pathway, multiple cytokines, prostaglandins, and hormones participate in it, such as IL-1, IL-6, vitamin D, and PTH [86]. Additionally, external factors could have a significant role in the regulation of RANKL, OPG, and even RANK to prevent bone loss or accelerate it. Agents like Zinc [87], probiotics [24], and exercise have shown to play a role in regulation [23].

CLA and RANKL/RANK/OPG

An examination was conducted in ovariectomized mice to clarify the effect of conjugated linoleic acid on postmenopausal bone conditions. This 24-week study compared the effects of safflower oil and CLA on bone inflammation, osteoblast genesis, and osteoclast genesis. The results indicated that using CLA could prevent RANKL expression, increase OPG expression, and regulate anti-inflammatory and pro-inflammatory cytokines [88].

To display the effect of various CLA isomers on agerelated bone resorption, a study was conducted on female mice. Sixty mice were split into four groups equally, and each group was fed corn oil, corn oil plus c9t11-CLA, corn oil plus t10c12-CLA, or corn oil plus t10c12-CLA plus c9t11-CLA. After six months, the results of the examinations demonstrated that sRANKL significantly decreased in groups fed t10c12-CLA and CLA-mix compared to other groups. Mice fed with t10c12-CLA showed higher bone mass density associated with a reduction in RANKL, TNFa, and IL-6 compared to c9t11-CLA [36].

Another in vitro study investigated the effect of CLA on human CD14+monocytes. This study showed that both c9t11 and t10c12 isomers of CLA decreased osteoclast proliferation and activity, but c9t11-CLA showed stronger results. Both of the isomers demonstrated decreasing effects on RANKL levels [89]. An experiment on murine RAW264.7 cells showed that CLA isomers prevented RANKL-induced osteoclastogenesis depending on their doses. CLA might reduce RANKL-induced TNF-a, and it could inhibit osteoclast-specific genes and osteoclast-specific transcription factors [90].

Glucosamine and RANKL/RANK/OPG

The results of a clinical trial indicated that OPG expression was unaffected by glucosamine sulfate, either alone or in combination with vitamin D3. Surprisingly, glucosamine enhanced RANKL levels both alone and in the presence of vitamin D3. All samples were derived from OA patients [35]. However, in the investigation by Yong Sun et al., glucosamine seemed to affect OPG and RANKL expression. In this study, conducted on Knee Osteoarthritis (KOA) patients, both the intervention and control groups took Etoricoxib as a basic medication, while the intervention group also received glucosamine. The reported outcomes indicated that OPG levels increased in both groups, particularly in the intervention group, and RANKL down-regulation was specifically observable in the glucosamine group [91].

Another study, performed on collagenase-induced osteoarthritis (CIOA) mice, demonstrated that glucosamine hydrochloride might reduce RANKL concentration. The outcome of this study shows that the amount of soluble RANKL in CIOA mice, after treatment with glucosamine hydrochloride, was significantly lower than in the non-treated group; however, it still was much higher than in healthy cases [92]. Igarashi et al.'s study on newborn mouse osteoblasts (Mc3T3-E1) concluded that glucosamine diminished RANKL expression. Based on the results of this study, glucosamine at a higher dosage (1mM) suppressed RANKL mRNA more effectively than at a lower dose (0.1mM) [93].

Conclusion

Bone health may be impacted by CLA and glucosamine supplements. Alongside direct anti-inflammatory mechanisms, they appear to influence the RANKL/RANK/OPG pathway to display their protective effects. More research is recommended to determine the impact of CLA and glucosamine supplements on bone health, with a focus on the RANKL/ RANK/OPG mechanism. Despite recent reports suggesting that using CLA could reduce RANKL expression, increase OPG expression, and regulate anti-inflammatory and proinflammatory cytokines, further investigation is necessary.

Authors' contributions Shaymaa J. Abdulrahman, Abduladheem Turki Jalil, Mohanad Ali Abdulhadi, Dumooa Falah, Muna S. Merza, Abbas F. Almulla, Ahmed Ali and Ronak Taher Ali all contributed significantly to the development and completion of this research paper. Shaymaa J. Abdulrahman, Abduladheem Turki Jalil and Mohanad Ali Abdulhadi were responsible for conceptualizing the study and designing the research framework. Ronak Taher Ali, Dumooa Falah, and Muna S. Merza conducted the literature review, gathering relevant information on bone health, osteoporosis, and the impact of various drugs and supplements. Abbas F. Almulla and Ahmed Ali were involved in analyzing and interpreting the data related to the regulatory effect of RANKL/RANK/OPG on bone turnover concerning the use of CLA and glucosamine supplements. All authors participated in the critical review of the manuscript, providing valuable insights, and revising the content to ensure the accuracy and clarity of the information presented. Shaymaa J. Abdulrahman, Abduladheem Turki Jalil, Mohanad Ali Abdulhadi, Dumooa Falah, Muna S. Merza, Abbas F. Almulla, Ahmed Ali and Ronak Taher Ali contributed equally in the preparation of the final version of the paper, approving it for submission, and agreeing to be accountable for all aspects of the work. Their collaborative efforts have been instrumental in addressing the knowledge gap regarding the potential effects of CLA and glucosamine on the RANKL/RANK/OPG mechanism on bone turnover, thus paving the way for the development of more effective treatment strategies and guiding future studies on these supplements and their mechanisms in bone health.

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Declarations

Competing interests The authors declare no competing interests.

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